Passion for Innovation. Compassion for Patients.™



Development of mRNA vaccines

Oct 5, 2021 Summary from DS seminar

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Vaccine Preparedness



To contribute to society through establishing pharmaceutical technology and manufacturing capability for vaccine preparedness

Stable vaccine supply through in-house manufacturing facility

- DS's marketed vaccines are being stably supplied from a domestic manufacturing facility
 - Seasonal influenza HA vaccine
 - Live vaccines (measles, rubella, and mumpus)



Vaccine R&D by utilizing innovative modality

Development of DS-5670*



- Initiated Ph1/2 study in March 2021
- DS-5670 utilizes original LNP that efficiently encapsulates mRNA and confers efficient delivery of mRNA to targets
- To build a platform that streamlines development and manufacturing of a variety of LNP-mRNA vaccines for future emerging/re-emerging infectious diseases

*Development of DS-5670 has been supported by AMED and MHLW

To build vaccine manufacturing facilities for future pandemics

- To establish in-house and domestic manufacturing facilities through an enterprise supported by MHLW
- To acquire capability of stable and emergency supply for vaccine preparedness and to become an essential infrastructure for emergency preparedness through collaboration with other organizations in the pharmaceutical industry



Agenda

1 LNP-mRNA vaccine technology

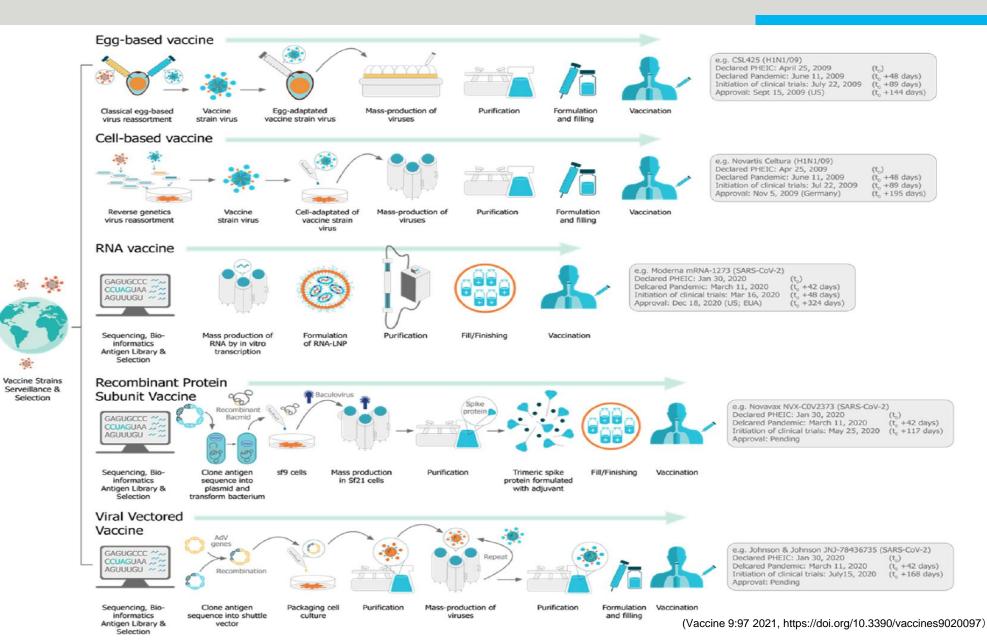
2 COVID-19 vaccine (DS-5670) preclinical data

3 Current status of DS-5670 development and future plan



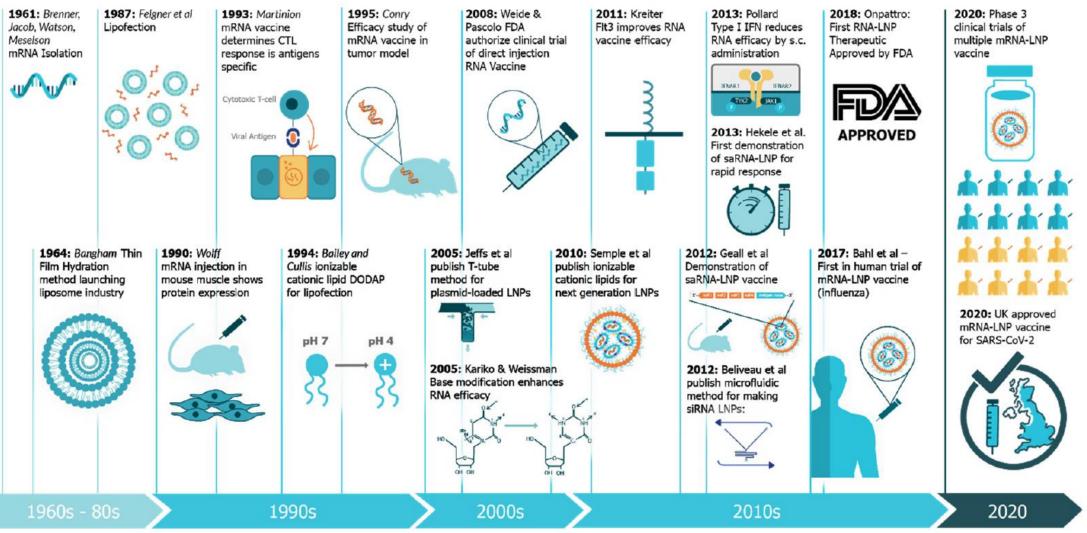
Manufacturing processes for different vaccine platforms





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History of technology development related to mRNA vaccines





The use of mRNA as a treatment modality (as of 2017)



| | No. of programs per R&D phase | | | | | | | |
|--|-------------------------------|-------|---|-----|---|----|-----|-------|
| mRNA modality | Res | Precl | 0 | IND | I | II | III | Total |
| Standardized cancer vaccines | | 1 | 1 | | 3 | 1 | | 6 |
| Individualized cancer vaccines | | | 1 | | 2 | | 1 | 4 |
| Therapeutic infectious disease vaccines | | | | | | 2 | | 2 |
| Prophylactic infectious disease vaccines & adjuvants | 4 | 4 | 1 | 1 | 6 | | | 16 |
| Replicon RNA infectious disease vaccines | 3 | | | | | | | 3 |
| Protein therapeutics for cancer & CV | | | 1 | 1 | 1 | | | 3 |
| Protein therapeutics for mono-genetic diseases | 8 | 8 | 3 | | | | | 19 |
| mRNA antibody therapeutics | 4 | 1 | | | | | | 5 |
| Ex vivo gene editing | 2 | | 2 | | | | | 4 |
| In vivo gene editing | 9 | 1 | | | | | | 10 |
| Ex vivo T cell engineering | | | 2 | | | | | 2 |

(mRNA Vaccines & Therapeutics 2017: an industry analysis of technologies, pipelines, stakeholders and deals released by La Merie Publishing on June 18, 2017)

Clinical studies assessing mRNA vaccines for infectious disease other than COVID-19 (as of Aug 2021)



| Disease target | Study stage | Delivery formulation | Status | Organization | |
|----------------------------|-------------|----------------------------------|-----------|---|--|
| CMV | Ph-2 | LNP | Ongoing | Moderna | |
| RSV | Ph-1 | Merck proprietary formulation | Ongoing | Merck/Moderna | |
| RSV | Ph-1 | Not disclosed | Completed | Merck/Moderna | |
| RSV | Ph-2 | LNP | Ongoing | Moderna | |
| Rabies | Ph-1 | Cationic lipid formulation | Ongoing | GSK | |
| Rabies | Ph-1 | LNP | Ongoing | CureVac | |
| Rabies | Ph-1 | Protamine | Completed | CureVac | |
| Chikungunya | Ph-1 | Not disclosed | Ongoing | Moderna | |
| hMPV/PIV3 | Ph-1 | LNP | Completed | Moderna | |
| Novel Flu (H10N8, H7N9) | Ph-1 | LNP | Completed | Moderna | |
| Zika | Ph-1 | LNP | Completed | Moderna | |
| Seasonal Flu | Ph-1 | LNP | Ongoing | Moderna, TranslateBio/SP, BioNTech/Phizer | |

mRNA vaccine candidates in clinical trials for COVID-19 (as of Sep 24, 2021)

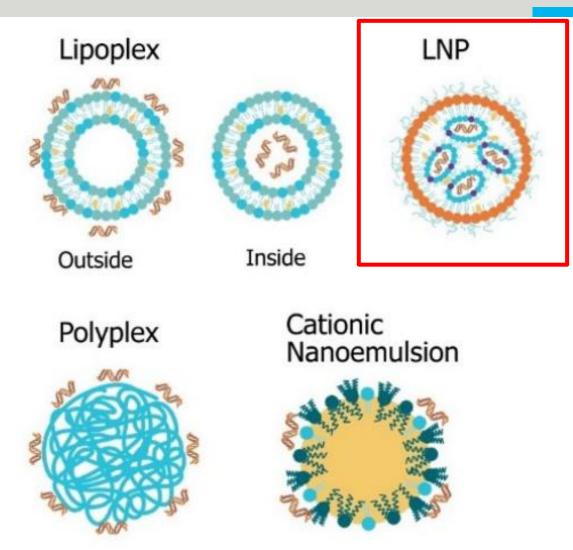


| ID ▼ | Vaccine platform acronym 🏹 | Vaccine platform description | Type of candidate vaccine | Number of doses | Schedule | Route of administratio | Developers | Phase |
|---------|----------------------------------|------------------------------|--|--------------------|----------------------------------|---------------------------|--|-----------|
| 9 | RNA | RNA based vaccine | mRNA-1273 | 2 | Day 0 + 28 | IM | Moderna + National Institute of Allergy and Infectious Diseases (NIAID) | Phase 4 |
| 10 | RNA | RNA based vaccine | BNT162b2 (3 LNP-mRNAs), also known as "Comirnaty" | 2 | Day 0 + 21 | IM | Pfizer/BioNTech + Fosun Pharma | Phase 4 |
| 12 | RNA | RNA based vaccine | CVnCoV Vaccine | 2 | Day 0 + 28 | IM | CureVac AG | Phase 3 |
| 22 | RNA | RNA based vaccine | ARCT-021 | NR | NR | IM | Arcturus Therapeutics | Phase 2 |
| 38 | RNA | RNA based vaccine | LNP-nCoVsaRNA | 2 | NR | IM | Imperial College London | Phase 1 |
| 39 | RNA | RNA based vaccine | SARS-CoV-2 mRNA vaccine (ARCoV) | 2 | Day 0 + 14 or Day 0 + 28 | IM | Academy of Military Science (AMS), Walvax Biotechnology and Suzhou Abogen Biosciences | Phase 3 |
| 46 | RNA | RNA based vaccine | ChulaCov19 mRNA vaccine | 2 | Day 0 + 21 | IM | Chulalongkorn University | Phase 1 |
| 71 | RNA | RNA based vaccine | PTX-COVID19-B, mRNA vaccine | 2 | Day 0 + 28 | IM | Providence Therapeutics | Phase 1 |
| 73 | RNA | RNA based vaccine | CoV2 SAM (LNP) vaccine. A self-amplifying mRNA (SAM) lipid nanoparticle (LNP) platform + Spike antigen | 2 | Day 0 + 30 | IM | GlaxoSmithKline | Phase 1 |
| 77 | RNA | RNA based vaccine | mRNA-1273.351. A lipid nanoparticle (LNP)-encapsulated mRNA-based vaccine that encodes for a full-length, prefusion | 3 | Day 0 or Day 0 + 28 or Day 56 | IM | Moderna + National Institute of Allergy and Infectious Diseases (NIAID) | Phase 4 |
| 82 | RNA | RNA based vaccine | MRT5500, an mRNA vaccine candidate | 2 | Day 0 + 21 | IM | Sanofi Pasteur and Translate Bio | Phase 2 |
| 85 | RNA | RNA based vaccine | DS-5670a, mRNA vaccine | 2 | NR | IM | Daiichi Sankyo Co., Ltd. | Phase 1/2 |
| 91 | RNA | RNA based vaccine | HDT-301: Self-replicating mRNA vaccine formulated as a lipid nanoparticle. | 2 | Day 0 + 28 | IM | SENAI CIMATEC | Phase 1 |
| 93 | RNA | RNA based vaccine | mRNA-1283 | 2 | Day 0 + 28 | IM | ModernaTX, Inc. | Phase 1 |
| 95 | RNA | RNA based vaccine | EXG-5003; a temperature-sensitive self-replicating RNA vaccine expressing the receptor binding domain of the SARS-CoV-2 spike protein. | 1 | Day 0 | ID | Elixirgen Therapeutics, Inc | Phase 1/2 |
| 98 | RNA | RNA based vaccine | mRNA COVID-19 vaccine | 2 | TBD | IM | Shanghai East Hospital and Stemirna Therapeutics | Phase 1 |
| 103 | RNA | RNA based vaccine | LNP-nCOV saRNA-02 vaccine; Self-amplifying RNA (saRNA) encapsulated in lipid nanoparticles (LNP) | 2 | Day 0 + 28 | IM | MRC/UVRI and LSHTM Uganda Research Unit | Phase 1 |
| 104 | RNA | RNA based vaccine | mRNA-1273.211. A multivalent booster candidate combining mRNA-1273 plus mRNA-1273.351. | 1 | Day 0 | IM | ModernaTX, Inc. | Phase 2/3 |
| 114 | RNA | RNA based vaccine | ARCT-154 mRNA Vaccine | 2 | Day 0 + 28 | IM | Arcturus Therapeutics, Inc. | Phase 2/3 |
| 115 | RNA | RNA based vaccine | ARCT-165 mRNA Vaccine | 2 | Day 0 + 29 | IM | Arcturus Therapeutics, Inc. | Phase 1/2 |
| 116 | RNA | RNA based vaccine | ARCT-021 mRNA Vaccine | 2 | Day 0 + 30 | IM | Arcturus Therapeutics, Inc. | Phase 1/2 |

(https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines)

Delivery systems for mRNA

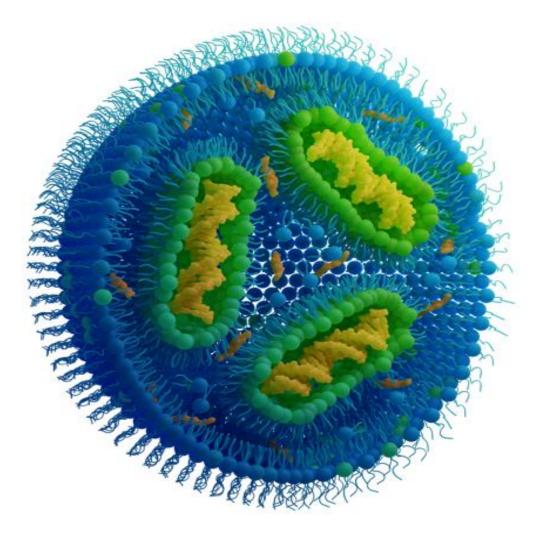




Non-viral mRNA delivery systems. Lipid-, polymer-, and emulsion-based delivery systems all use cationic groups to mediate condensation of the anionic RNA as well as delivery across the cell membrane.

Characteristics of DS's LNP-mRNA





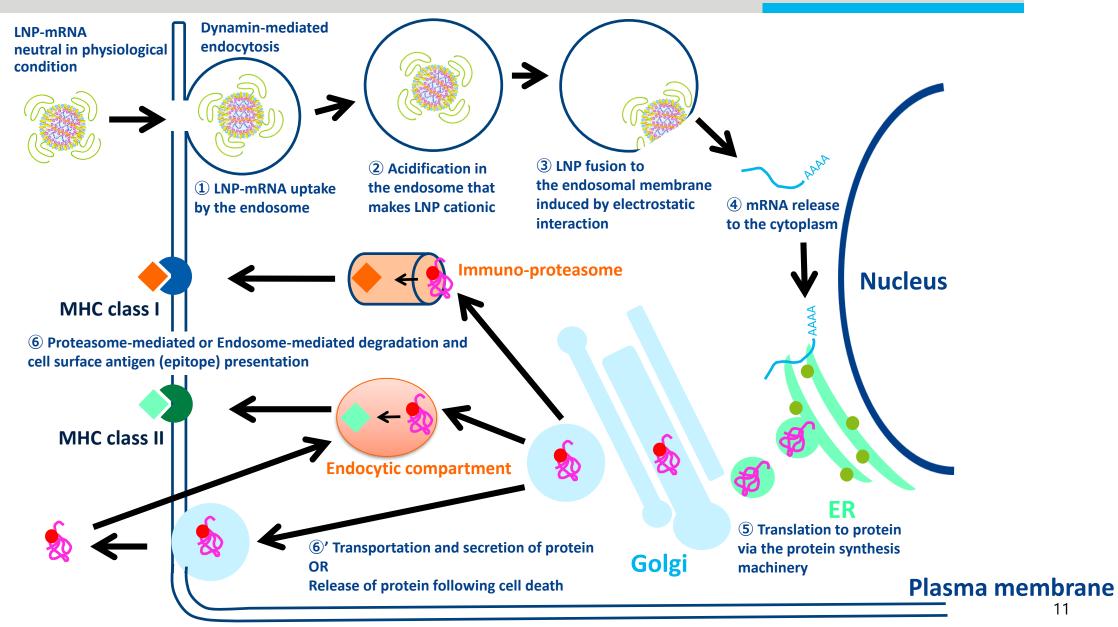
 DS original cationic lipid
Efficient encapsulation of mRNA in nanoparticles, and efficient delivery of mRNA to targets
Applicable to pandemic and other

vaccines

Proposed mechanism of LNP-mediated subcellular mRNA delivery and process of antigen protein

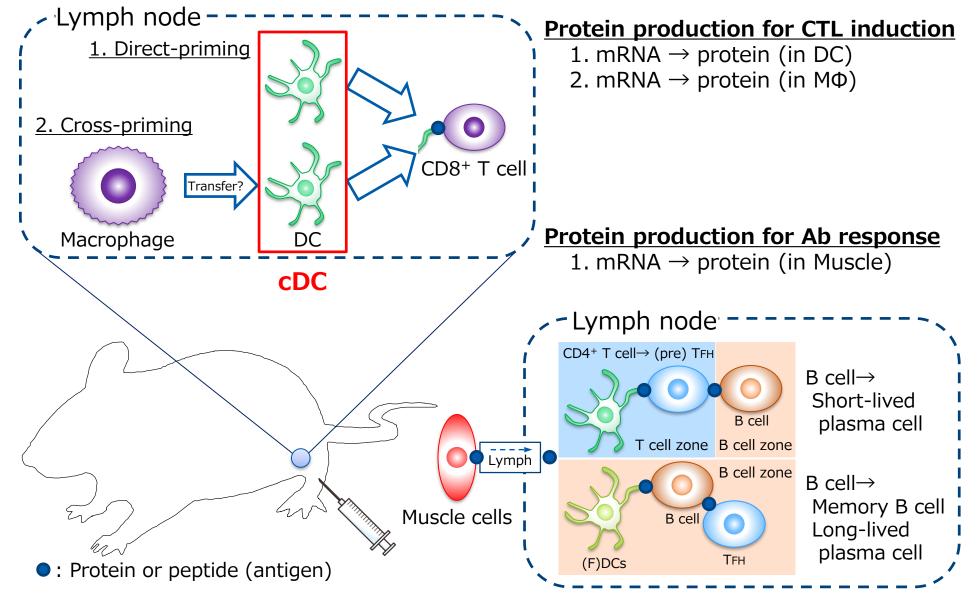


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Proposed immunogenic pathways of LNP-mRNA vaccine





CTL: cytotoxic T lymphocyte, DC: dendritic cells, MΦ: macrophage

Concept of LNP-mRNA vaccine (1/2)



[Pharmacological and safety profiles]

- 1. High-level, broad-spectrum antigen-specific immune responses are induced as compared with inactivated or recombinant protein antigens. In addition to antibody and helper T cell responses, cytotoxic T cells, which are necessary to eliminate intracellular pathogens, can be induced.
- 2. No interfering effect by existing immunity to vaccine formulations such as those observed in live attenuated vaccines and viral vectored vaccines confers stable boosting effects.
- 3. Due to high quality of antigen proteins produced in vivo, from the viewpoint of post-translational modification and conformation, induced immune responses are qualitatively superior to heterologously expressed antigen protein such as those produced in eggs, insects, or plants.
- 4. The risk of genetic injury in vaccines, such as carcinogenicity, immune deficiency, and transgenerational transmission, which poses a challenge to other types of genetic vaccines, is expected to be low.

Concept of LNP-mRNA vaccine (2/2)



[Quality and manufacturing profiles]

- 1. Lower risk in quality and manufacturing related to biologics, compared with live vaccines:
 - Non-pathogenic and relatively easy to handle in manufacturing
 - No requirement of bulky facilities for culture of cells or pathogens
 - The lack of in vivo replication ability makes it easier to determine dose
- 2. Once the platform has been established, development and manufacturing of a variety of LNP-mRNA vaccines can be streamlined

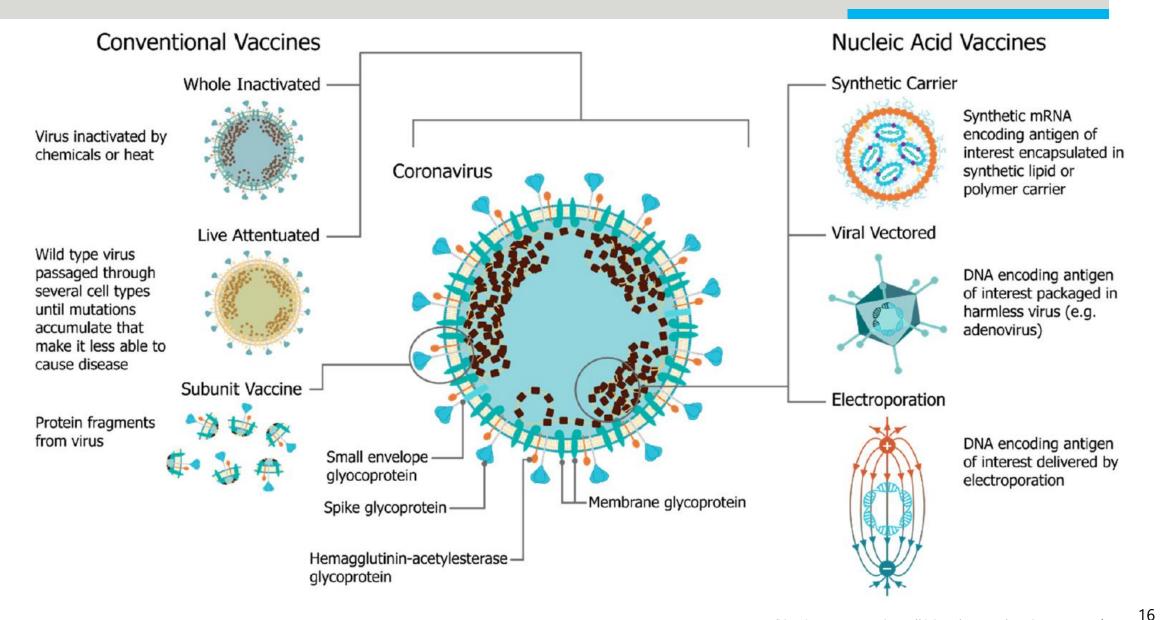
Significance of developing LNP-mRNA vaccines



- Original antigen design
- Feasible to supply vaccines containing novel antigens for breakthrough variants supposed to emerge in the future
- Having experiences in R&D for mRNA vaccine pipeline
- Expected to be superior in domestic development and distribution as compared with other leading mRNA vaccines developed in foreign countries
- To acquire capability of stable and emergency supply for vaccine preparedness and to become an essential infrastructure for emergency preparedness through collaboration with other organizations in the pharmaceutical industry

Structure of SARS-CoV-2 and vaccine modalities

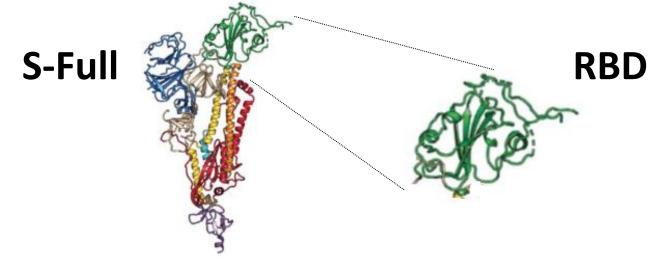




Design of SARS-CoV-2 spike protein (S) antigen for DS-5670

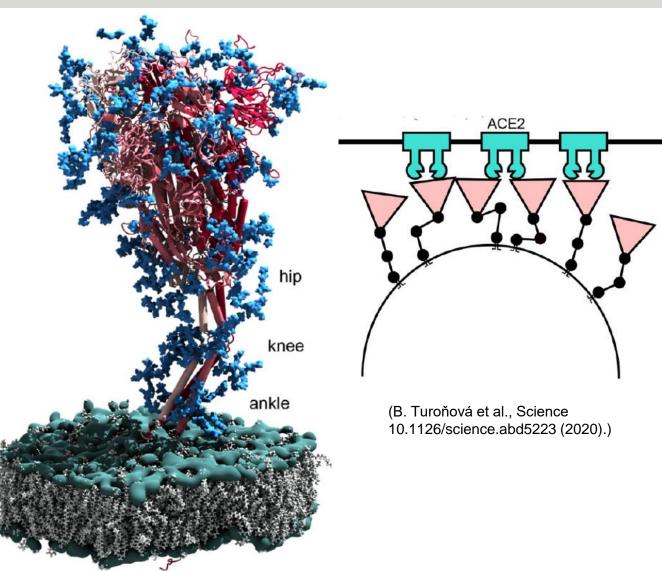


| Length of mRNA• 4.1 kb•Proposed advantages• May contain additional neutralization epitopes and T cell epitopes other than those present in RBD• | 1.0 kb |
|---|---|
| advantages epitopes and T cell epitopes other than | |
| | Efficient and stable encapsulation of mRNA into LNP because ORF of RBD is shorter than that of S-Full Lower risk of enhanced disease because potentially pathogenic epitopes are less as compared with S-Full (CELL 12060 https://doi.org/10.1016/j.cell.2021.05.032P NAS 117:8218 2020, Vaccine 25:2832 2007) |



Superiority of RBD antigen to S-Full antigen

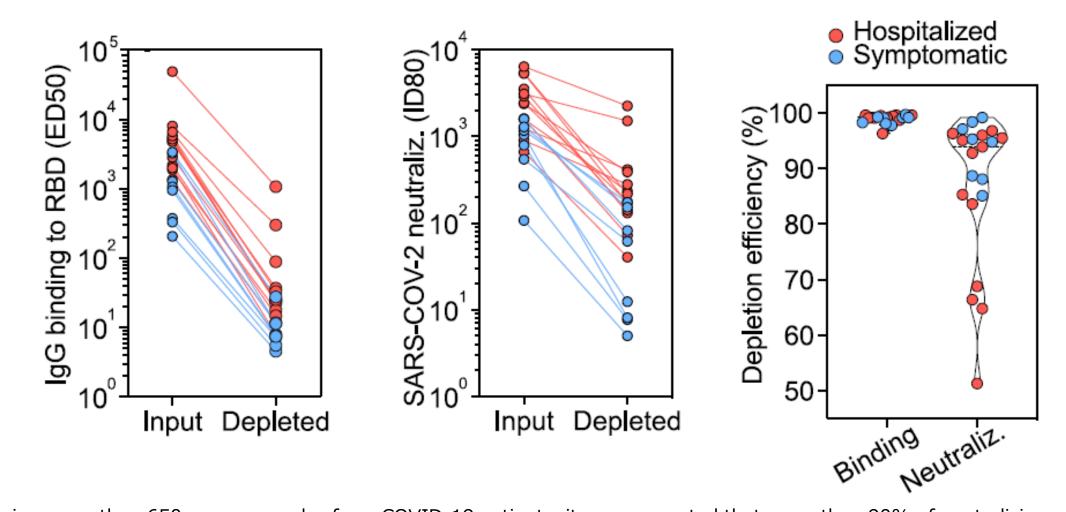




(B. Turoňová et al., Science 10.1126/science.abd5223 (2020).)

- Binding of RBD to ACE2 is cisregulated by domains other than RBD, so-called 'hip', 'knee', and 'ankle'.
- When using the S-Full of variants as vaccine antigen, mutations in 'hip', 'knee', and 'ankle' may affect the immunogenicity of RBD (may be evolutionally less immunogenic, enabling viral escape from host immune responses).
- In contrast, novel RBD antigens appropriate for emerging variants would be more simply designed and would be predictable.

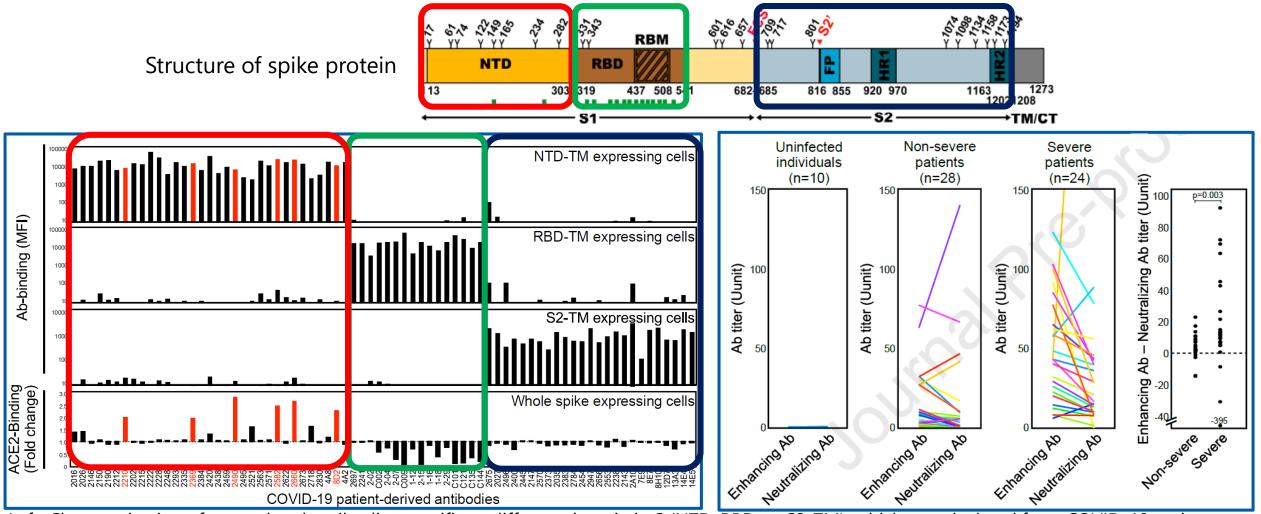
Critical role of RBD for inducing neutralizing activity



By analyzing more than 650 serum samples from COVID-19 patients, it was suggested that more than 90% of neutralizing antibodies targeted RBD

(The diagram shows the result of 21 samples, Cell 183:1024 2020)

Antibodies specific to N-terminal domain of spike protein and immune enhancement



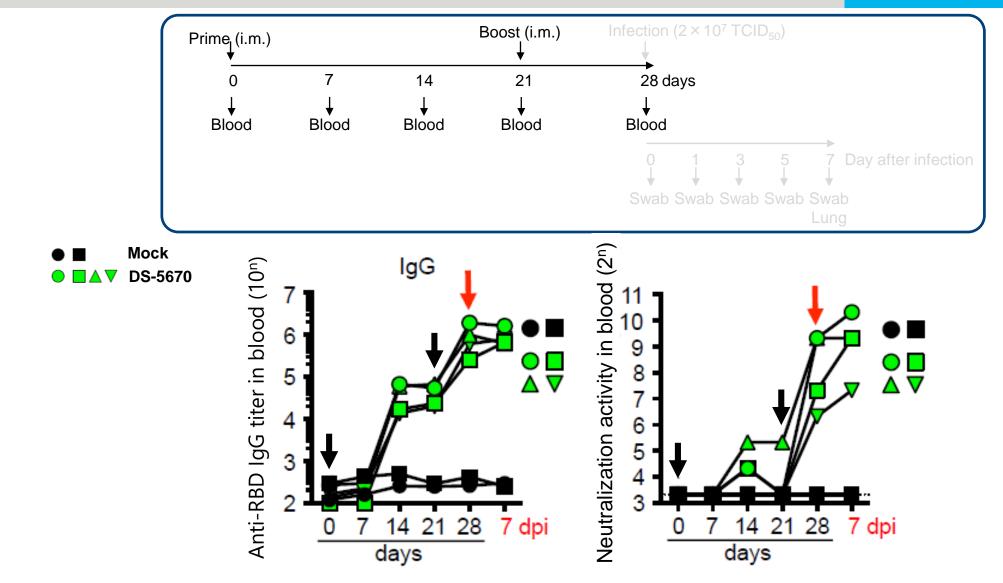
Left: Characterization of monoclonal antibodies specific to different domain in S (NTD, RBD, or S2-TM), which were isolated from COVID-19 patients. The lowest panel shows effects of monoclonal antibodies on S-Full binding to ACE2. Right: Enhanced and neutralizing antibody titers in serum obtained from COVID-19 patients.

Daiichi-Sankyo

Immunogenicity and protective efficacy of DS-5670 in cynomolgus monkeys (1/3)



Results of research collaboration with the University of Tokyo and Shiga Medical University*



*This data was acquired in the "Fundamental Research on the Control of a Novel Coronavirus (2019-nCoV), which is an initiative supported by the Japan Agency for Medical Research and Development (AMED). (Principal investigator: Prof. Yoshiro Kawaoka, Institute of Medical Sciences, The University of Tokyo 21

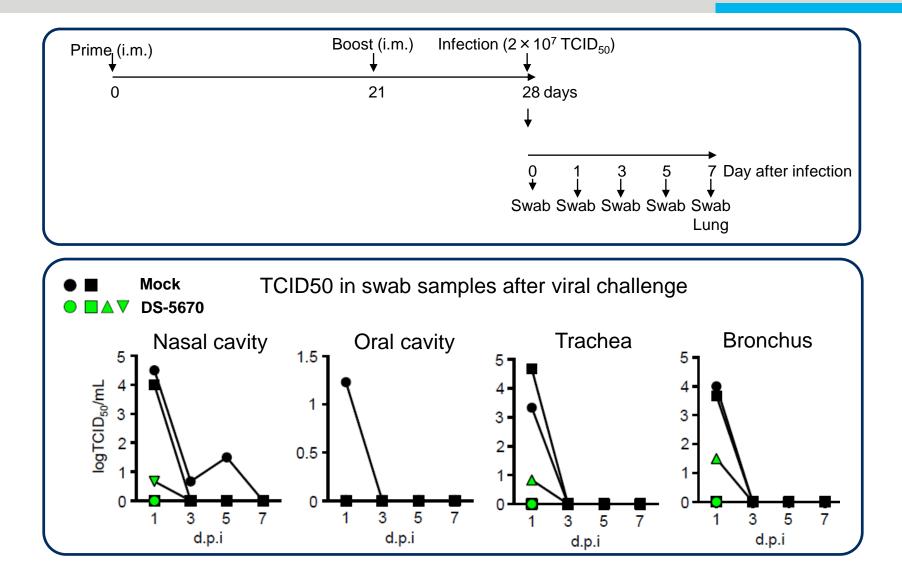
Immunogenicity and protective efficacy of DS-5670 in cynomolgus monkeys (2/3) Results of research collaboration with the University of Tokyo and Shiga Medical University* Daiichi-Sankyo

Oral cavity: IgG Nasal cavity: IgG Conjunctiva: IgG 5 5 5 anti-RBD titer (10ⁿ) anti-RBD titer (10ⁿ) anti-RBD titer (10ⁿ) 3 3 3 2 2 2 0 0 0 7 dpi 21 28 7 dpi 21 28 7 dpi 21 28 days days days Trachea: IgG Rectum: IgG Mock 5 5 anti-RBD titer (10ⁿ) anti-RBD titer (10ⁿ) DS-5670 4 3 3 2 2 0 21 28 7 dpi 28 7 dpi 21 days days

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Daiichi-Sankyo

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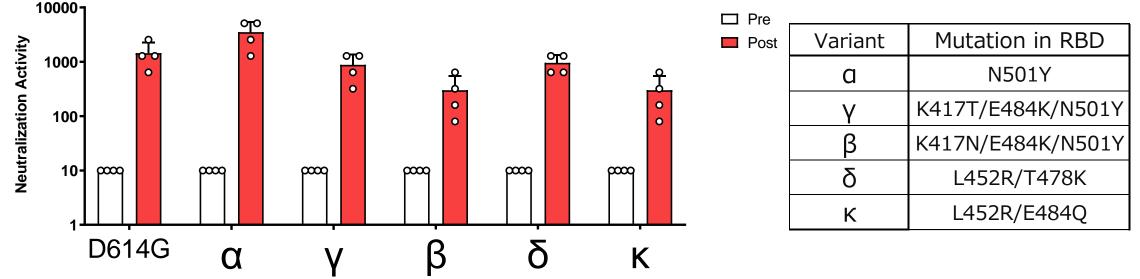


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Cross-neutralizing activity against recently emerged variants



- Cynomolgus monkey
- 50 μg/body of DS-5670 by mRNA conversion
- Dosed in brachial deltoid muscle q2w, total 3 times (4 monkeys/group)
- Measured neutralizing activity using plasma collected 2 weeks after the third dose (AMED Kawaoka group)



SARS-CoV-2 variant

| | SARS-CoV-2 variant | | | | | | | |
|-----------|--------------------|------|------|-----|------|-----|--|--|
| Monkey ID | D614G | a | γ | β | δ | к | | |
| #1 | 640 | 2560 | 640 | 160 | 1280 | 160 | | |
| #2 | 2560 | 5120 | 1280 | 640 | 640 | 640 | | |
| #3 | 1280 | 5120 | 1280 | 320 | 1280 | 320 | | |
| #4 | 1280 | 1280 | 320 | 80 | 640 | 80 | | |

*This data was acquired in the AMED's drug discovery support program "Development of a Vaccine for COVID-19 Vaccines".

Current status of DS-5670 development and future plan



- Selected to be a provider for the MHLW's "Emergent Initiative to Build Production Capacity for COVID-19 Vaccines^{*1} (First Round)"
- Selected to be a company for the AMED's drug discovery support program "Development of a Vaccine for COVID-19 Vaccines*² (Second Round)"
- Initiated Ph1/2 study in March 2021 and data expected around autumn 2021. Currently evaluating the safety, immunogenicity, and recommended dose.
- To initiate **active-controlled**, **non-inferiority confirmatory study** this year, in which several thousand subjects will be enrolled
- **BLA and commercialization expected in CY2022** when all regulatory requirements are satisfied
- A clinical trial for booster vaccination also being planned and considered
- The overall development plan and designs of further studies being continuously discussed with the Health Authority

The project aims to swiftly develop an actual (large-scale) production system for biologics, including vaccines, in order to ensure that the vaccines necessary for the prevention of the spread and severity of unexpected epidemics, including COVID-19, are produced as soon as possible, and that their supply is secured for the Japanese people. *2 The project aims to support the development of a vaccine against COVID-19, for which R&D is already underway, and aims to ensure the early commercialization of safe and effective vaccines.